

Independent Roles of Switching and Hypermutation in the Development and Persistence of B Lymphocyte Memory

Highlights

- The IgG1 BCR promotes the formation of bone-marrow plasma cells
- The IgG1 BCR disfavors the memory cell fate
- Somatic hypermutation reduces the longevity of memory B cells
- Polyreactive memory B cells are selected against over time

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In Brief

Activation-induced cytidine deaminase induces somatic hypermutation and class-switch recombination, which are essential to the development of humoral immunity. Nussenzweig and colleagues genetically separate the effects of these two diversification reactions and find that they have independent roles in the formation and persistence of memory in the antibody system.

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Independent Roles of Switching and Hypermutation in the Development and Persistence of B Lymphocyte Memory

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SUMMARY

Somatic hypermutation (SHM) and class-switch recombination (CSR) increase the affinity and diversify the effector functions of antibodies during immune responses. Although SHM and CSR are fundamentally different, their independent roles in regulating B cell fate have been difficult to uncouple because a single enzyme, activation-induced cytidine deaminase (encoded by *Aicda*), initiates both reactions. Here, we used a combination of *Aicda* and antibody mutant alleles that separate the effects of CSR and SHM on polyclonal immune responses. We found that class-switching to IgG1 biased the fate choice made by B cells, favoring the plasma cell over memory cell fate without significantly affecting clonal expansion in the germinal center (GC). In contrast, SHM reduced the longevity of memory B cells by creating polyreactive specificities that were selected against over time. Our data define the independent contributions of SHM and CSR to the generation and persistence of memory in the antibody system.

INTRODUCTION

During immune responses, B cells diversify their immunoglobulin genes in germinal centers (GCs) to produce the high affinity, class-switched antibodies that mediate humoral immunity (Allen et al., 2007a; Rajewsky, 1996; Victora and Nussenzweig, 2012). Antibody gene diversification is accomplished by somatic hypermutation (SHM) and class-switch recombination (CSR). Whereas SHM generates a pool of antibody variants with differing affinities, CSR exchanges the antibody constant region to produce antibodies with a diverse set of effector functions (Bournazos et al., 2015; Stavnezer et al., 2008). A single enzyme, activation-induced cytidine deaminase (AID), which is expressed primarily in the GC, initiates both SHM and CSR (Muramatsu et al., 2000). Although mutant forms of AID bias the reaction to SHM or CSR, the two diversification reactions are never completely separated (Barreto et al., 2003; Shinkura

et al., 2004; Ta et al., 2003; Wei et al., 2011). It has therefore been difficult to delineate the precise contributions of changes in affinity versus alterations in isotype to regulating the antibody response.

B cells expressing high affinity antibody variants are selectively expanded in the GC and preferentially seed the plasma cell compartment (Phan et al., 2006; Smith et al., 1997; Victora and Nussenzweig, 2012). As a result, serum antibody affinity increases over time, a phenomenon known as affinity maturation (Eisen and Siskind, 1964).

Although immunoglobulin E (IgE) expression is associated with limited bone-marrow plasma cell and memory B cell formation (He et al., 2013; Yang et al., 2012) and IgA expression promotes plasma cell differentiation (Duchez et al., 2010), the independent roles of SHM and IgG antibody class switching in regulating B cell fate are not well defined. Experiments using a transgenic IgG1 antigen receptor specific for hen egg lysozyme indicate that this isotype enhances clonal expansion and might bias B cells to become plasmablasts (Horikawa et al., 2007; Martin and Goodnow, 2002). However, an IgG1 BCR specific for 4-hydroxy-3-nitrophenylacetyl (NP) within the endogenous antibody locus fails to show the same effect (Kometani et al., 2013). Moreover, clonal analysis of wild-type and *Aicda*-deficient B cells that are unable to class switch showed that they were equally capable of producing plasmablasts (Taylor et al., 2015). Thus, whether the observed differences in B cell fate are due to antibody isotype, transgene-induced alterations in B cell development (Pogue and Goodnow, 2000; Roth et al., 1993; Roth et al., 1995), or differences in the antigen and/or affinity remains to be determined.

In addition to effects on B cell fate, differences in antibody isotype have also been associated with differences in the longevity of memory B cells (Pape et al., 2011). IgG⁺ memory B cells appear to have a far shorter half-life than IgM⁺ memory B cells. However, the antibodies expressed by IgG⁺ memory cells are more mutated than their IgM⁺ counterparts (Dogan et al., 2009; Pape et al., 2011). Thus, the relative contribution of hypermutation versus isotype switching to memory cell persistence remains to be determined.

Here, we report on experiments designed to uncouple CSR and SHM in order to examine their independent contributions to antigen-specific polyclonal B cell responses.

RESULTS

A Polyclonal System that Separates CSR and SHM

To examine the role of class switching during a polyclonal immune response, we immunized mice with R-phycoerytherin (PE) in complete Freund's adjuvant (CFA) (Pape et al., 2011). To track the fate of cells that had expressed *Aicda*, we combined a genetically targeted allele of *Aicda*, in which cre recombinase replaces the endogenous AID protein (*Aicda*^{cre/+}), with the *R26*^{LSL-YFP} allele, in which cre-mediated recombination leads to permanent yellow fluorescent protein (YFP) expression (Robbiani et al., 2008; Srinivas et al., 2001). Antigen-specific cells were enumerated by magnetic bead-based enrichment (Figure 2 and Figures S1A–S1C) (Pape et al., 2011). As expected, the majority of antigen-specific GC B cells and bone-marrow plasma cells (BMPCs) had expressed *Aicda* as determined by the YFP marker (90.5% and 83% YFP⁺, respectively), and most of these cells were class-switched (95.6% and 95.7%, respectively) (Figure S1B). In contrast, only 28.5% of antigen-specific memory cells were YFP⁺, of which only 48% were class-switched (Figure S1B).

To examine the contribution of class switching to the B cell response, we generated mice in which class switching takes place in the absence of AID and SHM (Figure 1A). To do so, we combined an *Aicda*-deficient allele (*Aicda*^{-/-}) with the aforementioned *Aicda*^{cre} allele (*Aicda*^{cre/-}) (Muramatsu et al., 2000). These *Aicda* mutant alleles were further crossed to the *R26*^{LSL-YFP} fate-mapping allele and to a genetically targeted Ig heavy chain (*Igh*) allele carrying *loxP* sites upstream of the μ and γ 1 constant regions (*Igh*^{96K}) (Bothmer et al., 2010). Activated B cells in the resulting *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice that transcribed the endogenous *Aicda* locus expressed cre in place of AID protein; cre expression recombines the *loxP* sites in the *Igh*^{96K} and *R26*^{LSL-YFP} loci thereby class switching the BCR to IgG1 while marking cells as YFP⁺ (Figure 1A). Because B cells in these mice do not express AID protein, they did not undergo SHM or suffer AID-induced DNA damage, thus allowing CSR to take place independently of SHM.

Flow cytometric analysis of Peyer's patches confirmed that GC B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice class switch robustly and exclusively to IgG1 (Figure 1B). The absence of class switching to IgA in Peyer's patches of *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice confirmed the absence of endogenous AID activity (Figure 1C).

The IgG1 BCR Mediates Selection into the BMPC Fate

Due to allelic exclusion, the vast majority of B cells express only one of their two *Igh* alleles (Cebra et al., 1966; Pernis et al., 1965). Thus, naive B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice expressed either the *Igh*^{96K} allele or the wild-type *Igh* allele (Figure S3A, see below). The ~50% of B cells with a productive V(D)J rearrangement in their *Igh*^{96K} allele could class switch to IgG1 upon cre expression; the other half of the B cell population, expressing the wild-type *Igh*, could not class switch because of the absence of both AID protein expression and *loxP* sites in the constant region of their expressed *Igh* gene.

After immunization, nearly all antigen-specific GC B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice became YFP⁺ due to high amounts of transcription of the *Aicda*^{cre} allele (Figures 2A–2C). Among GC B cells, ~31% were IgG1⁺ (Figures 2D and 2F). Taking

into consideration that only half of the B cells could class switch, this number of class-switched cells was similar to the frequency of class-switched GC cells in *Aicda*^{cre/+} *R26*^{LSL-YFP} mice immunized with PE (Figure S1). The antigen-specific memory pool contained 28% YFP⁺ cells in a resting state (Figures S1D and S2B), of which only 19% were class-switched, and this too was similar to *Aicda*^{cre/+} *R26*^{LSL-YFP} mice (Figures 2D and 2F and Figure S1). In contrast, antigen-specific BMPCs were 85% YFP⁺, of which nearly 79% of cells were class-switched (Figures 2E and 2F). Because only half of the responding B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice can class switch after activation (*Igh*^{96K}-expressers), no antigen-specific compartment should exceed ~50% class-switched unless the IgG1 BCR itself promotes such a fate. Thus, the finding that 79% of antigen-specific BMPCs were class-switched reveals IgG1-mediated positive selection into this compartment, which is independent of SHM.

IgG1 Alters GC LZ Gene Expression

To investigate differences in signaling by IgG1 and IgM BCRs that may account for the differential fate of these cells, we incorporated a Nurr77-GFP transgene that acts as a reporter of BCR signaling (Zikherman et al., 2012). Consistent with recent reports (Mueller et al., 2015), Nurr77 reporter expression was higher among light zone (LZ) cells than among dark zone (DZ) cells (Figure 3A). Moreover, transcriptional profiles between IgG1⁺ and IgM⁺ dark zone (DZ) cells were nearly indistinguishable ($r = 0.96$; Figure S2A), and a short pulse with the nucleotide analog 5-ethynyl-2'-deoxyuridine (EdU) revealed similar proliferation rates among IgM⁺ and IgG1⁺ GC cells (Figure S2B). In contrast, transcriptional profiles of IgG1⁺ and IgM⁺ LZ cells differed to a greater extent ($r = 0.86$; Figure S2A). A greater fraction of IgM⁺ LZ B cells were Nurr77⁺ compared to IgG1⁺ LZ B cells in *Aicda*^{cre/-} *Igh*^{96K/+} Nurr77-GFP mice (Figures 3B and 3C). We therefore purified and sequenced RNA from IgG1⁺ and IgM⁺ LZ cells that were Nurr77⁻ and Nurr77⁺ (Figure 3D).

Although we found little difference between Nurr77⁻ LZ cells of the two isotypes, Nurr77⁺ LZ cells differed in their transcriptional profiles to a far greater extent (Table S1 and Figure 3D), including differences in expression of chemokine receptors implicated in B cell migration in a manner that potentially presages the exit of IgG1⁺ cells from the GC and their differentiation into the BMPC fate (Figure 3D) (Cyster, 2003; Green and Cyster, 2012; Green et al., 2011; Hargreaves et al., 2001; Kabashima et al., 2006). For example, IgG1⁺ Nurr77⁺ cells had lower expression of CXCR5 and S1PR2 transcripts than their IgM⁺ counterparts (Figure 3D and Figures S2C and S2D), and expression of both of these receptors is essential to B cell retention within the follicle and the GC, respectively (Ansel et al., 2000; Förster et al., 1996; Green et al., 2011). In addition, expression of c-Myb, which has been implicated in CXCR4-mediated migration of plasma cells to the bone marrow (Good-Jacobson et al., 2015), was increased. We conclude that BCR signaling in the LZ differs in an isotype-specific manner, consistent with the biased fate choice made by IgG1⁺ B cells to the BMPC fate.

The IgG1 BCR Disfavors the Memory Cell Fate

Whereas IgG1⁺ cells constitute a majority of GC B cells and BMPCs, they represent a minority of the memory pool (Figures 2D and 2F and S1) (Dogana et al., 2009; Pape et al., 2011; Slifka

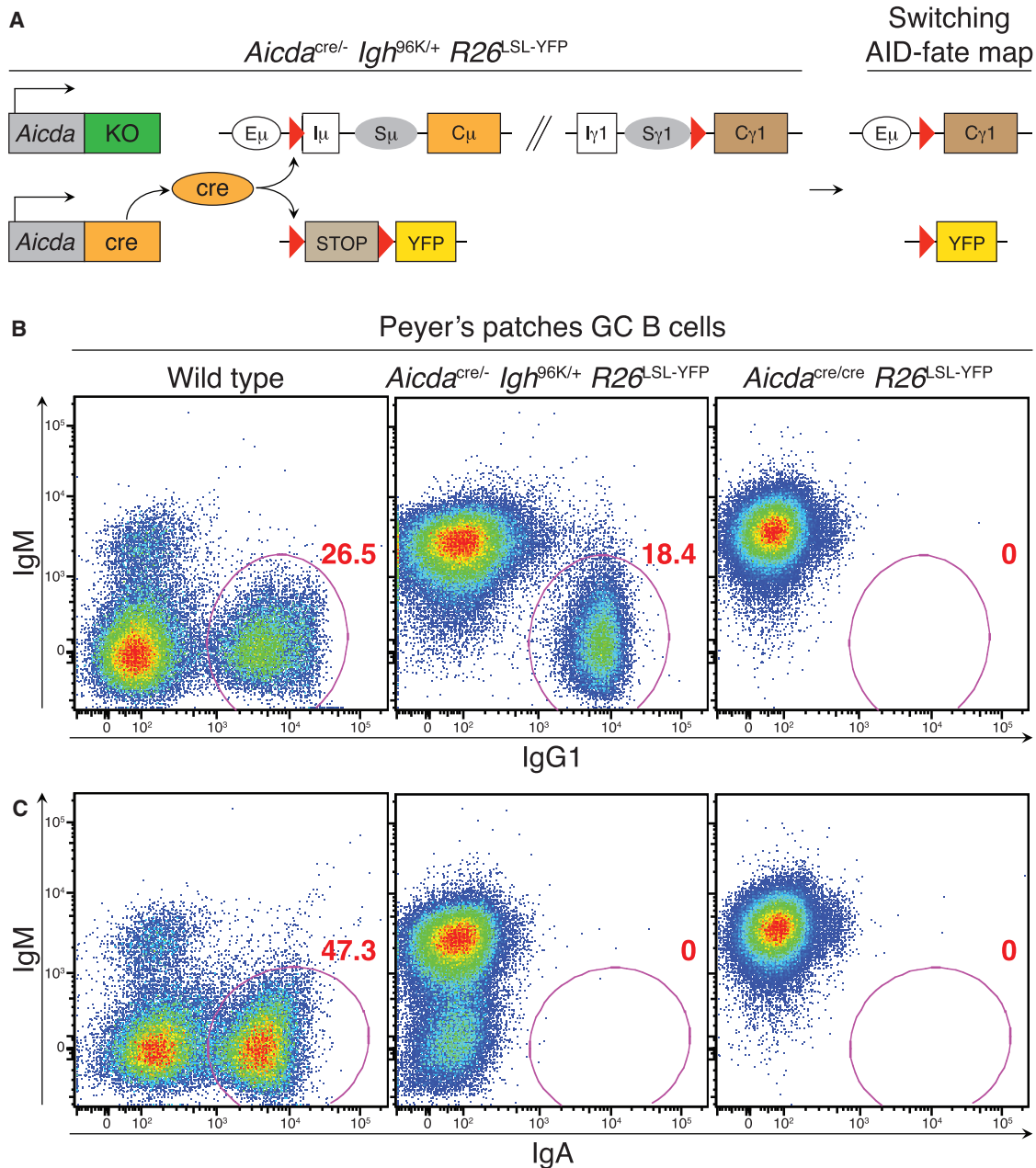


Figure 1. Uncoupling CSR and SHM in the Polyclonal System

(A) Diagram of allele combinations used to separate CSR and SHM. *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice express cre recombinase instead of AID protein, leading to cre-mediated class switching to IgG1 and fate-mapping based on *Aicda* expression in the absence of SHM.

(B and C) Class switching to IgG1 (B) and IgA (C) among all GC B cells in Peyer's patches of indicated mice. Results represent two independent experiments with at least two mice per genotype in each experiment.

et al., 1995). This raises the possibility that class switching biases responding B cells away from entering the memory pool. Alternatively, IgG1⁺ memory cells might not persist as long as IgM⁺ memory cells; however, this possibility was ruled out because once the memory pool was formed, the fraction of memory cells that were IgM and IgG1 remained stable over ≥ 1.5 years (Figure 5D and see below).

To test the notion that the IgG1 BCR counter-selects against the memory cell fate choice, we used *Igh* allotype-marked

mice in which the *Igh*^b allele was able to undergo isotype switching and the *Igh*^a allele was not (*Aicda*^{cre/cre} *Igh*^{96k(b)/WT(a)} *R26*^{LSL-YFP}; Figure 4A). In the absence of counter-selection, *Igh*^a- and *Igh*^b-expressing B cells should each comprise $\sim 50\%$ of the memory compartment (Figure S3A). However, we found that B cells expressing the *Igh*^b allele that can undergo class switching were under-represented in the memory compartment (Figures 4C–4E and S3). In contrast, control B cells carrying only a single *loxP* site in the *Igh*^b allele (Robbiani et al., 2008), and thus

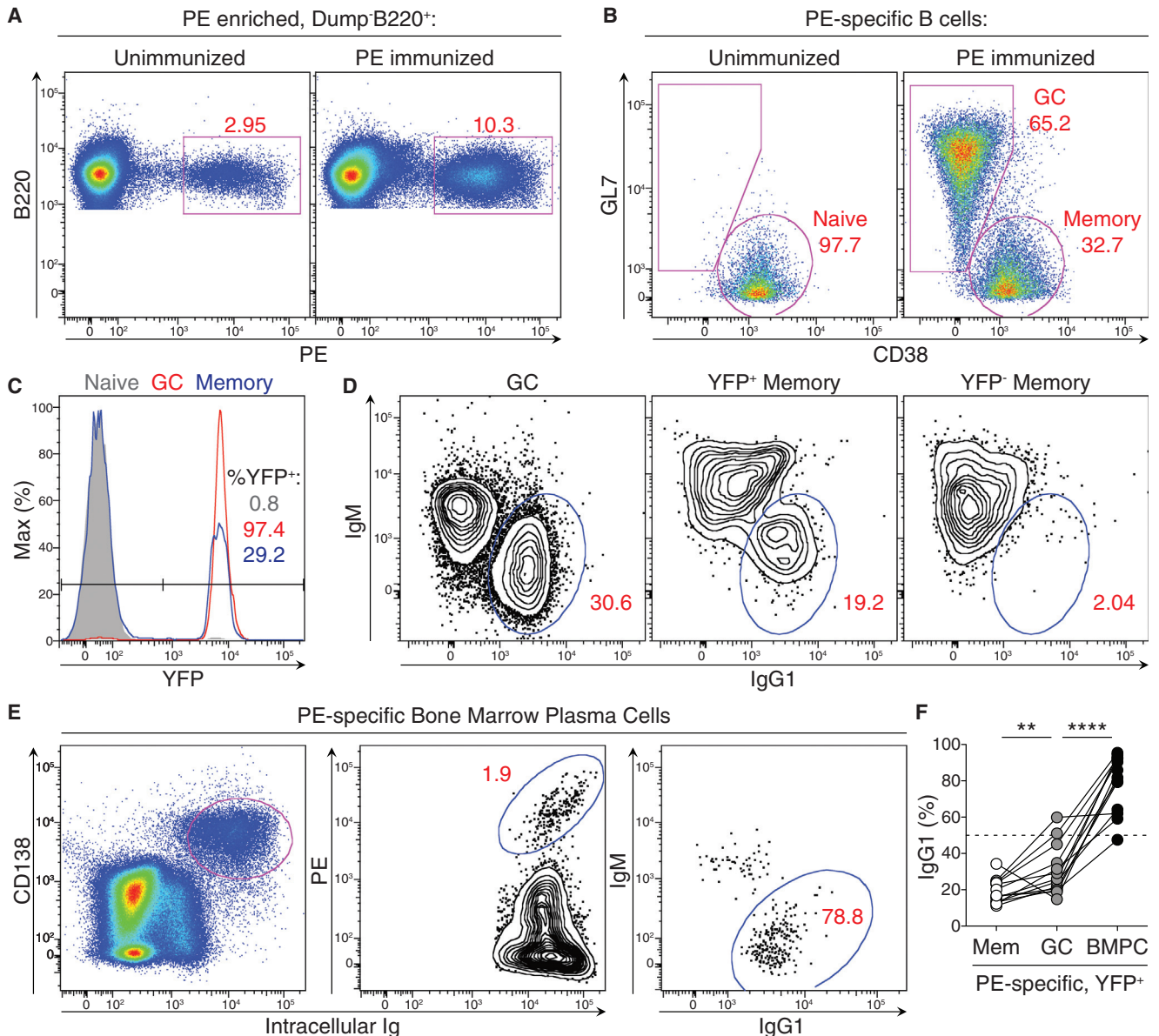


Figure 2. Antigen-Specific B Lineage Cells and Positive Selection for the Bone-Marrow Plasma Cell Fate

(A) PE-binding among Dump⁺ B220⁺ cells in naive and immunized *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice following column-based cell enrichment.

(B) GC and memory cells among antigen-specific B cells gated in (A) and identified based on surface expression of CD38 and GL7.

(C) Expression history of *Aicda* among antigen-specific naive B cells and GC and memory B cells, gated as in (A) and (B).

(D) IgM and IgG1 surface expression in indicated antigen-specific populations, gated as shown in (A)–(C).

(E) Plasma cells were enriched in immunized mice from the bone marrow based on CD138 surface expression and assessed for intracellular antigen-binding and isotype expression. BMPCs were further gated as Dump⁻YFP⁺.

(F) Fraction of IgG1⁺ cells among YFP⁺ cells in the antigen-specific memory, GC, and BMPC compartments of *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice 40–59 days after immunization with PE. Results are combined from three independent experiments; each line connects the indicated cell populations from the same animal. ****p < 0.0001, **p = 0.009. Two-tailed Mann-Whitney test was used to assess significance. See also Figure S1.

unable to class switch (*Aicda*^{cre/cre} *Igh*^{μ-1(b)/WT(a)} *R26*^{LSL-YFP}; Figure 4B), were not counter-selected in the memory compartment (Figures 4C–4E and S3).

To corroborate these findings in an independent system, and to examine the kinetics of counter-selection of IgG1 memory cells, we generated competitive bone-marrow chimeras. In

control chimeras, neither of the two *Igh* alleles were able to class switch (50% *Aicda*^{cre} *R26*^{LSL-YFP}; 50% *Aicda*^{cre} *Igh*^{μ-1} *R26*^{LSL-YFP}); neither of these B cell subsets expressed AID protein, and *Igh*^{μ-1} could not undergo cre-mediated class switching. In experimental chimeras, 50% of the cells were able to class switch to IgG1 (50% *Aicda*^{cre} *Igh*^{96K} *R26*^{LSL-YFP}; 50% *Aicda*^{cre}

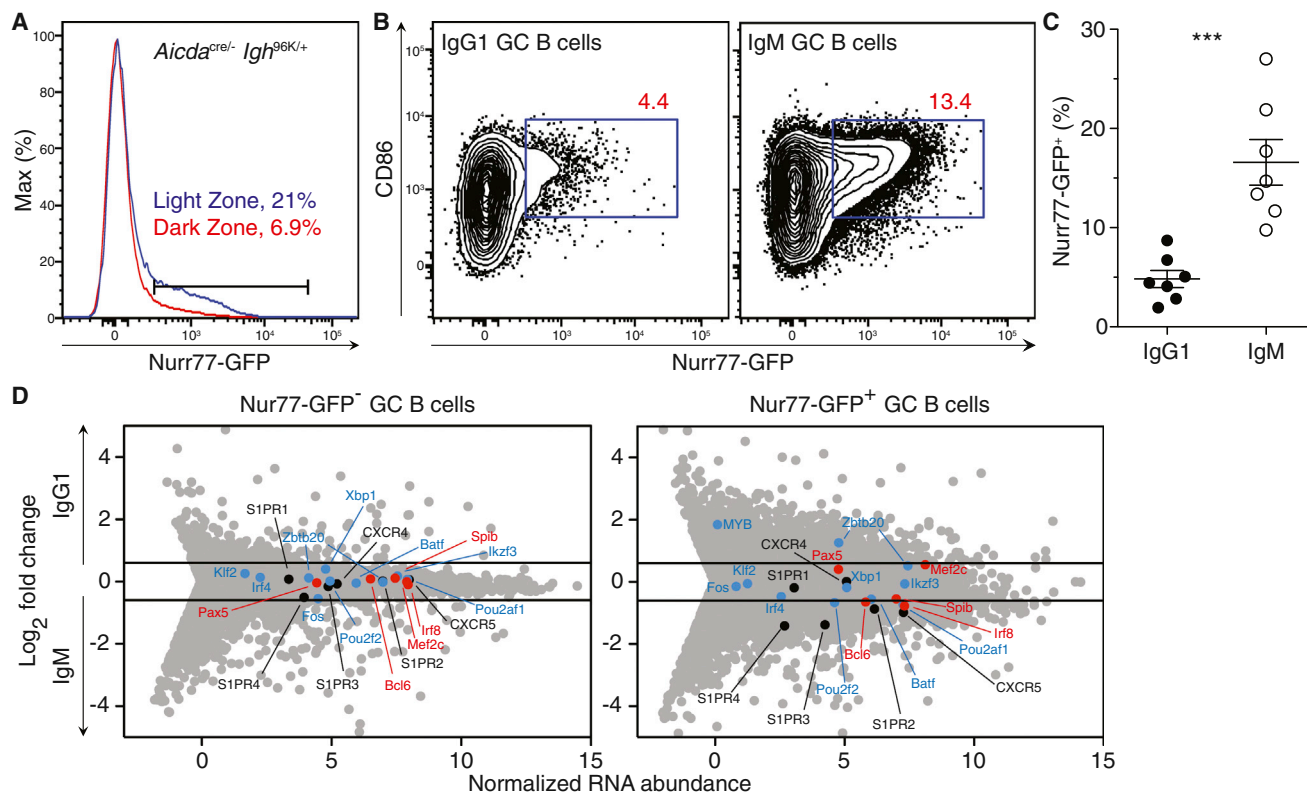


Figure 3. Isotype-Specific Signaling in the GC

(A) Histogram showing Nurr77-GFP expression among light zone ($CXCR4^{lo}CD86^{hi}$) and dark zone ($CXCR4^{hi}CD86^{lo}$) GC B cells 32–41 days after immunization with PE.

(B) Representative flow cytometry plots showing Nurr77-GFP expression among IgM^{+} and $IgG1^{+}$ GC B cells.

(C) Fraction of Nurr77-GFP⁺ cells among IgM^{+} and $IgG1^{+}$ GC B cells, gated as shown in (B). Line indicates mean.

(D) RNA sequencing analysis showing selected chemokine receptor genes (black) and negative (red) and positive (blue) regulators of plasma cell differentiation (see [Experimental Procedures](#)) among IgM^{+} and $IgG1^{+}$ LZ cells that were Nurr77⁻ (left plot) and Nurr77⁺ (right plot). Lines represent cut-offs for genes up- or downregulated by a fold-change of at least 0.6 (\log_2). Results represent two independent experiments for (A)–(D) with $n = 3$ –5 mice per genotype for each experiment. Error bars represent SEM. *** $p = 0.0006$. Two-tailed Mann-Whitney test was used to assess significance. See also [Figure S2](#) and [Table S1](#).

$Igh^{hi-1} R26^{LSL-YFP}$), and counter-selection against $IgG1^{+}$ B cells would result in a decrease in the frequency of $Aicda^{cre} Igh^{96K} R26^{LSL-YFP}$ B cells ($CD45.2^{+}$) in the memory pool.

Chimeras were immunized with PE in CFA and antigen-specific B cells were enumerated over time. On day 10 after immunization, cells that were and were not able to undergo CSR were equally represented in the GC and YFP⁺ memory compartment ([Figure 4F](#)). In contrast, by day 32, when memory and GC populations had undergone significant expansion, B cells able to class switch became under-represented in the memory pool but not in the GC, an effect that persisted even 70 days after immunization ([Figure 4F](#)). Notably, counter-selection became most prominent between 20 and 32 days after immunization, excluding a major contribution by the extrafollicular B cell response, which resolves earlier ([Jacob et al., 1991; Liu et al., 1991; Taylor et al., 2012b](#)). We conclude that, independent of SHM or AID expression, the class-switched BCR biases the fate choice made by B cells entering the long-lived compartment of B lineage cells.

AID Expression Reduces the Longevity of Memory B Cells

The persistence of memory cells is essential for humoral immunity ([McHeyzer-Williams et al., 2012; Tarlinton and Good-Jacobson, 2013](#)). Given that IgM^{+} memory cells that have relatively few mutations persist longer than IgG^{+} memory cells that have a higher load of hypermutation ([Pape et al., 2011](#)), we asked whether SHM or CSR have a role in memory cell persistence.

To address this question, we tracked long-term memory B cell responses in two groups of mice: (1) $Aicda^{cre/+} R26^{LSL-YFP}$ mice, in which activated B cells that express AID protein are marked by YFP expression; (2) $Aicda^{cre/-} Igh^{96K/+} R26^{LSL-YFP}$ mice, in which B cells could not express AID protein but could nevertheless undergo CSR to $IgG1$ and were marked by YFP expression when they expressed cre from the *Aicda* locus.

GC responses in both sets of mice showed similar kinetics, peaking between 2 and 5 weeks after immunization and subsiding by 125 days ([Figure 5A](#)). In addition, nearly all antigen-specific $IgG1^{+}$ memory cells in both groups of mice were YFP⁺. In contrast, only 16% and 22% of IgM^{+} memory cells were YFP⁺

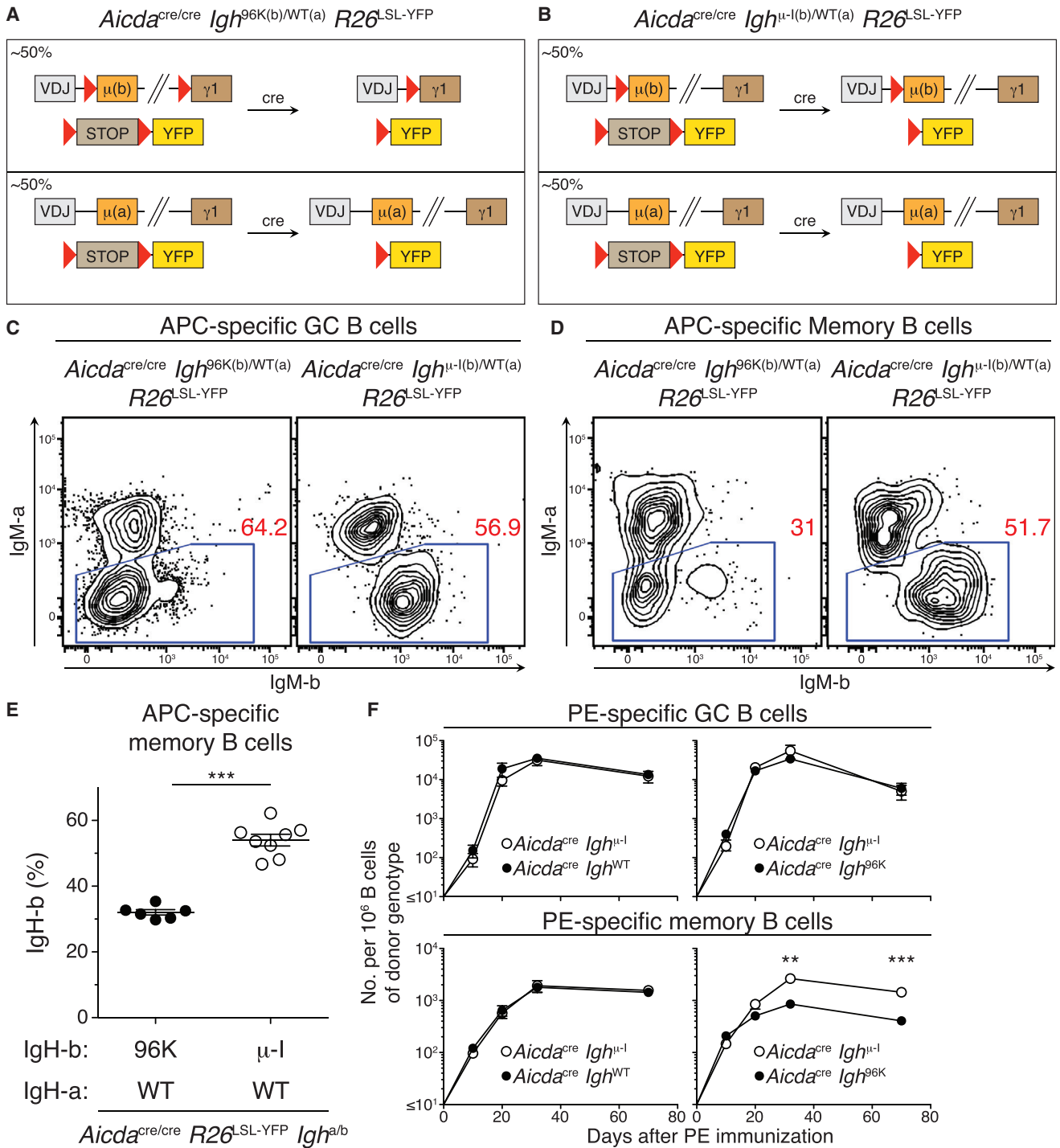


Figure 4. Counter-Selection by IgG1 BCR for Memory Cell Fate

(A and B) Diagrams of allele combinations used to assess IgG1-mediated counter-selection in the memory cell fate in (C)–(E).

(C and D) IgM^a and IgM^b were examined by flow cytometry 21 days after immunization with APC in CFA in the antigen-specific GC (C) and memory (D) compartments of indicated mice.

(E) Fraction of antigen-specific YFP⁺ memory B cells expressing the *Igh*^b allotype in indicated mice 40 days after immunization with APC in CFA. Lines represent means.

(F) Mean number of antigen-specific GC (upper panels) and YFP⁺ memory (lower panels) B cells of indicated genotype per 10⁶ total B cells of the donor genotype in bone-marrow chimeras immunized with PE in CFA. Results represent three independent experiments with a total of 8–10 mice of each genotype in (C) and (D); two independent experiments with a total of 6–8 mice total of each genotype in (E); and two independent experiments with a total of 6–7 mice per group at each time point in (F). Error bars represent SEM. **p = 0.0022; ***p ≤ 0.0007. Two-tailed Mann-Whitney test was used to assess significance. See also Figure S3.

in *Aicda*^{cre/+} *R26*^{LSL-YFP} and *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP}, respectively (Figure S4A). The majority of IgM⁺ YFP⁻ memory B cells were bona fide memory cells because they were significantly expanded in number compared with naive animals (Figure S4B). However, it remains possible that a minor subset of this population may not have responded to the immunization.

Consistent with previous observations (Pape et al., 2011), IgM⁺ memory cells that had not expressed AID (YFP⁻), which represent most of the IgM⁺ memory population, did not appreciably decline for 562 days after immunization in both sets of mice (Figure S4C). In contrast, AID-experienced (YFP⁺) memory cells in *Aicda*^{cre/+} *R26*^{LSL-YFP} mice underwent a long-term decline (Figures 5B and 5C). As noted earlier, the fraction of IgM⁺ and IgG1⁺ cells within the surviving AID-experienced memory population did not change over time, indicating that the negative effect of AID on memory cell persistence was independent of BCR isotype (Figure 5D). Furthermore, a history of AID protein expression per se contributed to the decline of memory cells because YFP-marked IgM⁺ and IgG1⁺ memory cells generated in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice, which transcribed the *Aicda* locus but did not express AID protein, persisted at far higher frequencies throughout the period of observation (Figures 5B and 5C and Figure S4D). Even after GC responses had dissipated in both sets of mice (day 125; Figure 5A), YFP⁺ memory cells continued to decline at a faster rate in *Aicda*^{cre/+} *R26*^{LSL-YFP} mice than in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice (Figure S4E). Thus, differences in GC kinetics could not account for the differences in memory cell longevity observed. In contrast to the memory cells, antigen-specific BMPCs of both isotypes persisted similarly in both genotypes of mice (Figures 5E and 5F). We conclude that AID expression negatively affects the persistence of memory B cells in a manner that is independent of class switching, suggesting that SHM might alter memory B cell longevity.

Memory B Cells with Polyreactive BCRs Have Diminished Longevity

A large proportion of nascent B cells carry polyreactive antibodies, but this reactivity is removed from the repertoire at two distinct checkpoints during development (Wardemann et al., 2003). During the GC response, up to 40% of B cells reacquire polyreactivity by SHM (Koelsch et al., 2007; Mouquet et al., 2010; Tiller et al., 2007). To determine whether the reacquisition of polyreactivity accounts for the long-term decline of AID-experienced memory B cells, we cloned antibodies from single antigen-specific IgG1⁺ memory B cells at early (day 31) and late (day 347⁺) time points after immunization. Early and late memory antibodies that bound to the cognate antigen (PE; Figure 6A) were tested for polyreactivity. In healthy humans, a substantial fraction of non-specific IgG⁺ memory B cells are polyreactive (Tiller et al., 2007). Consistent with these findings, ~33% of antigen-specific early memory antibodies were highly polyreactive (Figures 6A and 6B). In contrast, late memory B cells carried antibodies that were devoid of such reactivity (Figures 6A and 6B; $p = 0.0102$). As might be expected for cells that acquired polyreactivity, Nurr77-GFP expression was significantly higher among IgG1⁺ memory B cells that had expressed AID than among those generated in the absence of AID (Figures 6C and 6D). We conclude that AID expression is associated with

increased polyreactivity and that this reactivity is removed from the memory compartment over time.

DISCUSSION

Humoral immune memory is essential for vaccines and protection against re-infection. Memory depends on B cell clonal expansion, differentiation, and long-term persistence of antigen-specific cells (McHeyzer-Williams et al., 2012; Rajewsky, 1996; Tarlinton and Good-Jacobson, 2013; Taylor et al., 2012a). Despite their importance, the roles of SHM and CSR in governing the development and longevity of memory in the antibody system are not well defined. Here we have addressed these issues by genetically separating SHM and CSR, which normally occur in parallel, thereby revealing their independent contributions to humoral immune memory.

The antibody response includes two waves of B cell differentiation. The extrafollicular B cell response generates an early wave of plasmablasts that are short-lived, and memory B cells that produce germline-encoded or sparsely-mutated antibodies. Concomitantly, the GC response forms and persists for up to several weeks after the extrafollicular response subsides (Victoria and Nussenzweig, 2012). As a result, the GC supplies an essential source of somatically hypermutated and class-switched B lineage cells that enter the memory and plasma cell pools.

Within the GC, somatic antibody mutants are iteratively selected in an affinity-dependent manner. GC selection is mediated by BCR-dependent uptake of antigen for presentation as peptide-MHCII to CD4⁺ T follicular helper cells (Allen et al., 2007b; Shulman et al., 2014; Victoria et al., 2010). High-affinity clones thereby receive greater T cell help and consequently reside for a longer period of time in the DZ where they undergo accelerated cell cycles, allowing such clones to expand within the population (Gitlin et al., 2015; Gitlin et al., 2014). Although the BCR plays a key role in this process, by capturing and internalizing antigen, the contribution of BCR isotype and signaling to GC dynamics and output remain to be defined.

A multitude of signals can induce isotype-switching, including those derived from cytokines, Toll-like receptor ligands, CD40 activation, and BCR cross-linking (Stavnezer et al., 2008). Moreover, high-affinity B cells might preferentially elicit many of these signals. Distinguishing the effects of these signal(s) and affinity-enhancing mutations, both of which might precede and lead to class switching, from the effect of the class-switched BCR itself has been difficult. For example, the BMPC compartment is enriched in both affinity-enhancing mutations and class-switched cells (Slifka et al., 1995; Smith et al., 1997). Thus, whether class-switching simply occurred *en passant* with affinity-enhancing mutations and other signal(s) that mediated selection into the BMPC pool, or whether the class-switched BCR per se played a deterministic role in BMPC formation, remained unclear.

Using a polyclonal system in which class-switching is mediated by cre recombinase in the absence of SHM, we found that class switching to IgG1 is sufficient to bias B cell fate choice into the long-lived compartment of effector cells without affecting expansion in the GC. It is possible that differences in germline variable gene usage could emerge among *Igh*^{96K}-expressing and *Igh*^{WT}-expressing B cells in *Aicda*^{cre/-} *Igh*^{96K/+}

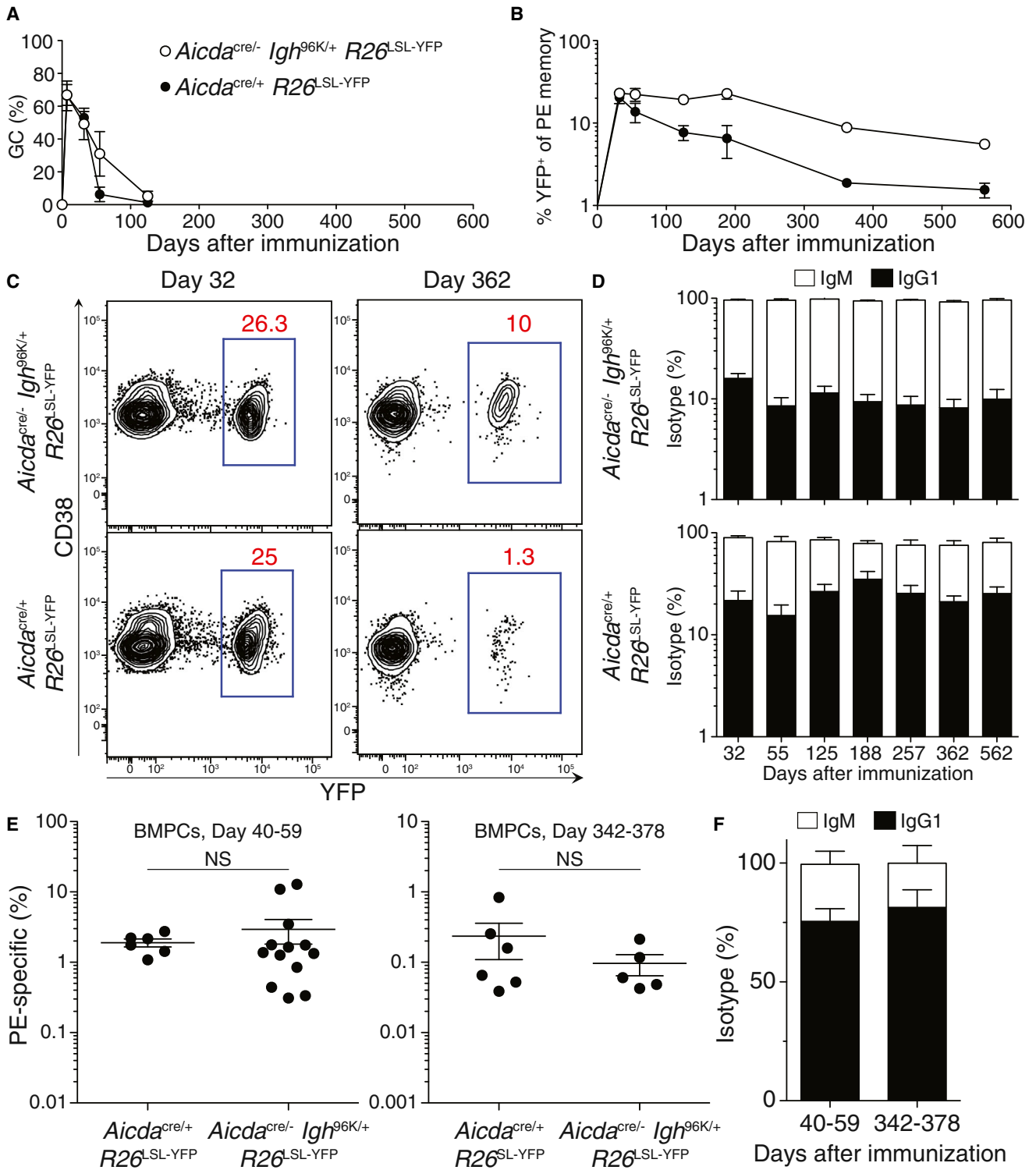


Figure 5. Role of CSR and SHM in Memory and Plasma Cell Longevity

(A) Mean fraction of GC cells among antigen-specific B cells in indicated mice after immunization with PE in CFA.

(B) Mean fraction of YFP⁺ cells among antigen-specific memory B cells in indicated mice after immunization with PE in CFA.

(C) Representative flow cytometry plots showing YFP expression among antigen-specific memory B cells in indicated mice at 32 and 362 days after immunization.

(D) Mean fraction of IgG1⁺ and IgM⁺ cells within the PE-specific YFP⁺ memory cell compartment in the indicated mice after immunization.

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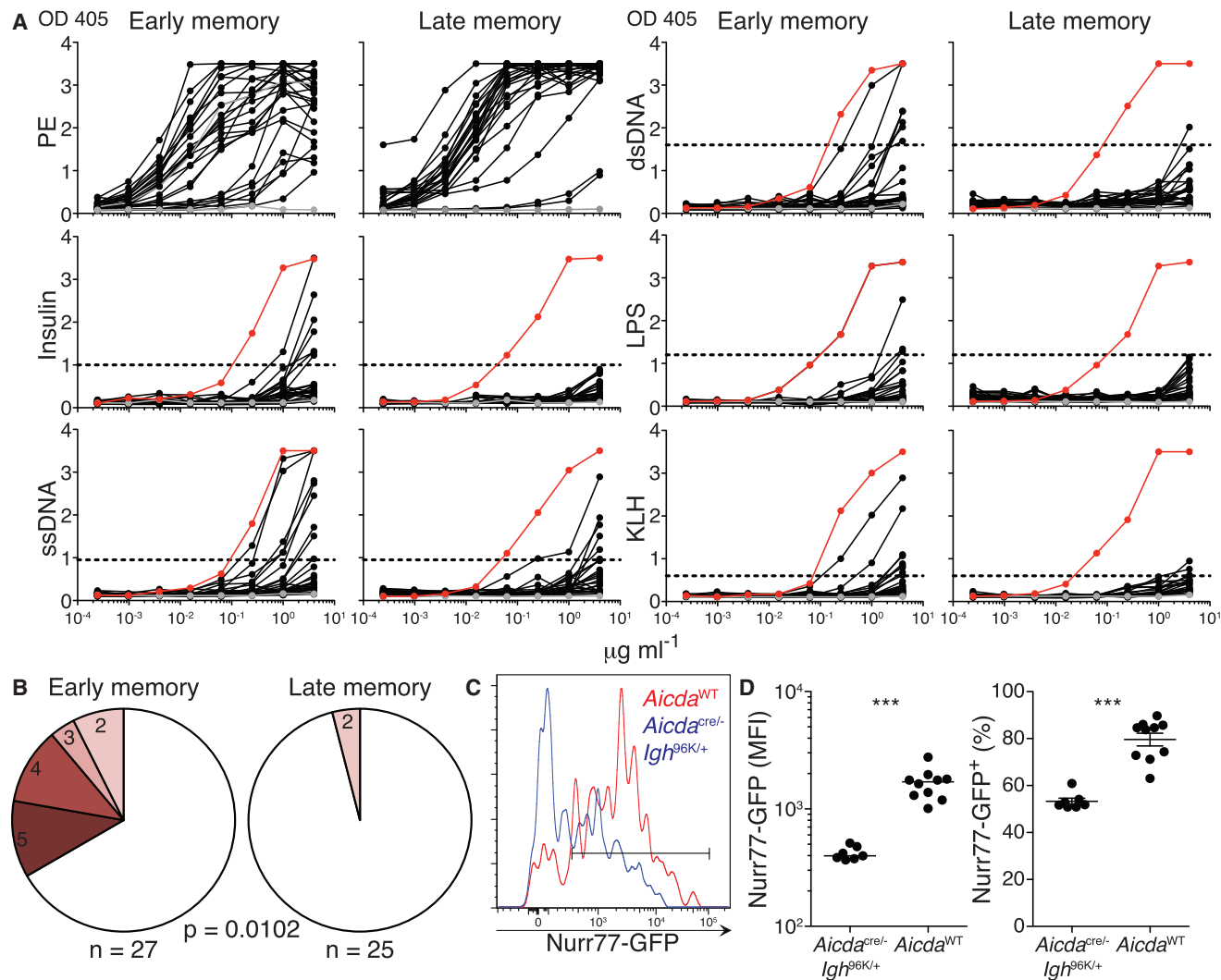


Figure 6. Selection against Memory Cell Polyreactivity

(A) ELISAs measuring binding of early and late memory antibodies to PE, insulin, ssDNA, dsDNA, LPS, and KLH. Horizontal lines show cut-off values for positive reactivity. Grey lines represent negative control antibody mGO53; red lines represent positive control antibody ED38 (Meffre et al., 2004).

(B) Pie charts of early and late memory antibodies showing fraction of antibodies that react with 2, 3, 4, or all 5 antigens tested in polyreactivity ELISAs. p value compares fraction of antibodies that bind to ≥ 3 structurally distinct antigens.

(C) Nurr77-GFP expression among PE-specific IgG1 memory B cells of indicated genotypes at 32–42 days after immunization.

(D) Median fluorescence intensity and fraction of Nurr77-GFP⁺ cells among PE-specific IgG1 memory B cells. Results are from two independent experiments. Three to five mice per genotype were used in each experiment in (C) and (D). Each dot represents one animal and line indicates median (left graph) or mean (right graph) in (D). Error bars represent SEM. *** $p \leq 0.0007$. Fisher's exact test and two-tailed Mann-Whitney test were used to assess statistical significance in (B) and (D), respectively. See also Figure S5.

R26^{LSL-YFP} mice. However, any such differences would necessarily arise due to the altered isotype of *Igh*^{96K}-expressing B cells, because no other differences exist between these two types of allelically excluded B cells within the same mouse. Moreover, *Igh*^{96K}-expressing B cells are unlikely to selectively

utilize higher affinity germline-variable genes, because these B cells had similar profiles of cognate antigen-binding in the GC and did not outcompete B cells that did not class switch.

Whereas class-switched B cells were positively selected into the BMPC pool, they were counter-selected from the memory

(E) Frequency of PE-specific cells in the BMPC compartment of the indicated mice at early (days 40–59) or late (days 342–378) time points after immunization. Lines represent means.

(F) Mean fraction IgG1⁺ and IgM⁺ cells among PE-specific BMPCs at the indicated time points after immunization of *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice. Results are from two to three independent experiments for each genotype and at each time point. Error bars represent SEM. Each dot represents one animal in (E). Significance was assessed using a two-tailed Mann-Whitney test. Error bars represent SEM. See also Figure S4.

compartment. Once the memory and BMPC pools had formed, the fractions of IgM and IgG1 cells within these compartments were stable for ≥ 1 year, suggesting that the class-switched BCR affects entry into, rather than persistence within, these compartments. The role of isotype in fate choice is likely to have physiologic relevance since the antibodies encoded by isotype-switched B cells possess unique Fc-mediated effector functions that are particularly well-suited for the antibody-secreting compartment of BMPCs. In contrast, IgM⁺ B cells might be better suited as a source of memory cells that can re-diversify their antibodies and subsequently class switch during secondary responses to antigenically related pathogens (Dogan et al., 2009; Pape et al., 2011; Purtha et al., 2011).

In addition to altering the membrane external constant region, isotype switching alters the cytoplasmic tail of the BCR in an evolutionarily conserved and isotype-specific manner (Achatz et al., 1997; Kaisho et al., 1997; Reth, 1992). In vitro experiments indicate that the IgG cytoplasmic tail alters the signal transducing effector molecules that associate with the BCR (Engels et al., 2009; Engels et al., 2014; Liu et al., 2012; Wakabayashi et al., 2002), increases BCR clustering (Liu et al., 2010), and crosslinking-induced Ca²⁺ responses (Horikawa et al., 2007; Waisman et al., 2007). We found that the IgG1 BCR differentially regulated signaling in the GC LZ as revealed by altered Nur77-GFP expression and chemokine receptor expression, suggesting a mechanism for isotype-specific B cell exit from the GC and preferential entry into the BMPC fate. Consistent with the idea that isotype switching per se can alter B cell fate, IgE expression has been associated with shortened residence in the GC and rapid differentiation into short-lived plasma cells, without prominent contribution to the memory or BMPC pools (He et al., 2013; Yang et al., 2012). Moreover, transgenic IgA expression has also been described to promote plasma cell differentiation (Duchez et al., 2010). Thus, isotype-specific BCR signaling appears to generally contribute to GC B cell fate decisions, in a manner unique to each isotype.

The maintenance of serum antibody varies considerably among different vaccines (Amanna et al., 2007). However, characterizing memory B cells has been challenging, especially in human studies that frequently rely on in vitro techniques to enumerate such cells by eliciting antibody secretion (Amanna et al., 2007; Crotty et al., 2004; Crotty et al., 2003). Nevertheless, the available data indicates that $\sim 10\%$ of the memory cell population survives 10 years after immunization and is then stably maintained (Crotty et al., 2003; Taylor et al., 2012a; Yu et al., 2008).

In mice, the persistence of memory B cells is independent of both helper T cells (Vieira and Rajewsky, 1990) and the persistence of cognate antigen (Anderson et al., 2006; Maruyama et al., 2000). However, longevity varies among memory B cells expressing different isotypes: IgM⁺ memory B cells, which as a group have a lesser amount of SHM, were found to be longer-lived than IgG⁺ memory B cells, which are more mutated, raising the possibility that memory cell persistence might be controlled by BCR isotype and/or SHM (Pape et al., 2011). We found that the rate of memory B cell decline was dependent on AID expression but not on CSR. Superficially, our findings might appear to be at odds with those of Jenkins and colleagues, but in fact they are entirely in keeping with their report of a difference in

longevity between IgM⁺ and IgG⁺ memory B cells because IgG⁺ cells are uniformly AID-experienced while only a fraction of IgM⁺ memory B cells have expressed high amounts of AID (Dogan et al., 2009; Pape et al., 2011). More importantly, our experiments provide a mechanistic explanation for the observation that memory B cells that have expressed high amounts of AID have a shorter half-life, thereby revealing a previously undocumented tolerance checkpoint in the antibody system.

The nascent B cell repertoire contains a substantial fraction of antibodies with poly- and self-reactivity, both of which are removed at two distinct checkpoints during B cell maturation (Wardemann et al., 2003). However, up to 40% of antibodies regain these features by SHM in the GC (Tiller et al., 2007). Although the most highly self-reactive of these antibodies are eliminated in the GC upon encountering autoantigen (Chan et al., 2012), polyreactive antibody mutants frequently enter the memory pool (Koelsch et al., 2007; Mouquet et al., 2010; Tiller et al., 2007). Moreover, SHM is a frequent source of autoantibodies in diseases such as systemic lupus erythematosus (Mietzner et al., 2008). Therefore, while a checkpoint against autoreactivity exists in the GC, it appears to eliminate only the most autoreactive antibodies and/or GC-expressed autoantigens (Chan et al., 2012; Pulendran et al., 1995; Shokat and Goodnow, 1995). Notably, mature B cells that are transgenically engineered to encounter a high concentration of autoantigen in the periphery are rapidly eliminated by clonal deletion (Lam and Rajewsky, 1998; Ota et al., 2011). Whether autoreactive specificities that enter the memory pool are culled by a similar mechanism remains to be determined. We conclude that differential persistence of autoreactive antibody mutants in the memory compartment represents a tolerance checkpoint in the antibody system.

EXPERIMENTAL PROCEDURES

Mice

Aicda^{cre}, *Aicda*^{-/-}, *R26*^{LSL-YFP}, *Igh*^{hi-1}, and *Igh*^{96K} were described previously (Bothmer et al., 2010; Muramatsu et al., 2000; Robbiani et al., 2008; Srinivas et al., 2001). Nur77-GFP mice were obtained from A. Weiss (Zikherman et al., 2012). B6.SJL mice and B6.Cg-Gpi1a Thy1a Igha/J were obtained from Jackson Laboratories. Bone-marrow chimeras were made for experiments in Figure 4F and for experiments involving Nur77-GFP transgenic cells in Figure 3B-D and Figure 6C and 6D. To generate chimeras, we irradiated C57BL/6 or B6.SJL males with two doses of 525 rad 3 hr apart. 5–10 $\times 10^6$ bone-marrow cells from donor genotypes were injected into recipients, which were analyzed at least 8 weeks later. All experiments were performed with authorization from the Institutional Review Board and the IACUC at The Rockefeller University.

Cell Enrichments

Spleen and lymph node cells were purified by forcing tissue through 40 μ m mesh into complete RPMI media (GIBCO) containing 6% serum. PE-specific B cells were enriched as previously described (Pape et al., 2011; Taylor et al., 2012b). To enrich plasma cells, we made cell suspensions from bone marrow in PBS containing 1 mM EDTA and 2% serum. Anti-CD138-APC (BD Biosciences) and anti-APC microbeads (Miltenyi Biotec) were used for enrichment with magnetized LS columns. In Figure 3, APC-specific cells were enriched by negative selection using anti-CD4-biotin, anti-CD8-biotin, anti-Gr1-biotin, and anti-Ter119-biotin (eBioscience), streptavidin microbeads (Miltenyi Biotec), and magnetized LS columns. Following column-based enrichment, samples were processed for flow cytometry. Cell counts from column-bound and flow-through samples were assessed using AccuCheck counting beads (Invitrogen) and cell numbers from both fractions were evaluated.

Immunizations and Treatments

Mice were immunized with 15 μg of PE or cross-linked APC (Prozyme) in a 50 μl emulsion with CFA (Sigma) at a single subcutaneous site at the base of the tail. To examine proliferating cells, we injected mice with 1 mg EdU (Life Technologies) in PBS 2 hr before analysis. Cells were then stained for flow cytometry and processed using a Click-iT EdU-Pacific Blue kit (Life Technologies) according to manufacturer's instructions.

Flow Cytometry and Cell Sorting

Single cell suspensions were processed in PBS that was maintained at 4°C and contained 2% serum and 1 mM EDTA. Samples were treated at 4°C for 10 min with 1 $\mu\text{g}/\text{ml}$ of anti-CD16/32 (2.4G2, Bio-X-Cell) and then stained for 25 min at 4°C. Antibodies used for flow cytometry were from eBioscience, BD Biosciences, Biolegend, and Life Technologies. Streptavidin-Qdot800 was from Life Technologies and Streptavidin-PerCP-Cy5.5 was from eBioscience. Cell fixation and permeabilization were performed using the Cytofix/Cytoperm kit (BD Biosciences). Samples were analyzed on a BD Fortessa. GC B cells were gated as live, single, and Dump(CD4, CD8, Gr-1, F4/80)⁻B220⁺CD38⁻GL7⁺ and PE binding. Memory B cells were gated as live, single, and Dump⁻B220⁺CD38⁺GL7⁻ and PE binding (Pape et al., 2011; Ridderstad and Tarlinton, 1998; Taylor et al., 2012b). DZ and LZ surface phenotype was assessed using CXCR4 and CD86, as described previously (Victoria et al., 2010). BMPCs were gated as live, single, and Dump⁻CD138⁺ and R26^{YFP+} and/or intracellular Ig^{high}. Antigen-specific BMPCs were identified by surface and intracellular staining with either PE or APC. CD45.1 and CD45.2 allotypic markers were used to trace donor genotypes in bone-marrow chimera experiments. *Aicda*^{cre} R26^{LSL-YFP} and *Aicda*^{cre} *Igh*^{96K} R26^{LSL-YFP} cells were CD45.2⁺, and *Aicda*^{cre} *Igh* ^{μ -1} R26^{LSL-YFP} cells were CD45.1⁺. For RNA sequencing, cells were sorted directly into Trizol LS reagent (Invitrogen) that was maintained at 4°C using a FACS Aria II (Becton Dickinson). For antibody cloning, memory B cells were single-cell sorted into 96 well plates using a FACS Aria II (Becton Dickinson). Antibodies from sorted cells were cloned and produced as described previously (Dosenovic et al., 2015).

Gene-Expression Analysis

Purified RNA from sorted samples was processed using the SMARTer Ultra Low Input RNA for Illumina Sequencing kit (Clontech Laboratories). High throughput sequencing was performed using a HiSeq 2500 (Illumina). In Figure 3D, genes referred to as positive and negative regulators of plasma cell differentiation were curated based on the literature (Nutt et al., 2015). Positive regulators included *Irf4*, *Zbtb20*, *Pou2f1*, *Pou2f2*, *Ikzf3*, *Batf*, *Fos*, *Klf2*, and *Xbp1*. Negative regulators included *Irf8*, *Bcl6*, *Pax5*, *Spib*, and *Mef2c*. Chemokine receptors included *CXCR4*, *CXCR5*, *S1PR1*, *S1PR2*, *S1PR3* and *S1PR4*. A SYBR Green based PCR mix (ThermoFisher Scientific) was used for quantitative PCR analysis. RNA-seq reads were aligned with STAR version 2.3.0 allowing unique alignments and using Mouse Ensembl genes as reference. Differential expression was calculated using Cufflinks with default settings.

ELISA

ELISA for PE binding was performed by overnight coating of high binding 96-well plates (Corning Incorporated) with 50 μl of 4 $\mu\text{g}/\text{ml}$ PE in PBS. Plates were then washed with PBS containing 0.05% TWEEN 20 (Sigma) and then blocked with PBS containing 1% BSA. Monoclonal antibodies were incubated for 1 hr at indicated dilutions in PBS. Plates were then washed and incubated with HRP conjugated anti-mouse IgG (Jackson Immunoresearch). Polyreactivity was assessed using LPS from *E. coli* (Sigma), human insulin (Sigma), dsDNA (Sigma) and ssDNA (prepared from dsDNA by heating at 95°C for 30 min), and KLH (Sigma) (Mouquet et al., 2010). High binding plates were incubated overnight with 50 μl of ssDNA, dsDNA, LPS, and KLH at 10 $\mu\text{g}/\text{ml}$ each and insulin at 5 $\mu\text{g}/\text{ml}$. Plates were then washed with PBS containing 0.001% TWEEN 20. Plates were blocked for 2 hr at room temperature in PBS containing 1 mM EDTA, 0.05% TWEEN 20 and 2.5% bovine serum albumin (Sigma). After washing, monoclonal antibodies were incubated for 2 hr at indicated dilutions in PBS. Plates were then washed and incubated with HRP conjugated anti-mouse IgG (Jackson Immunoresearch) for 1 hr. After washing, plates were developed by adding HRP substrate (Life Technologies). Absorbance was measured at 405 nm.

Statistical Analyses

Statistical significance was assessed using the tests indicated in the figures using Prism software v. 5.0 (Graphpad).

ACCESSION NUMBERS

The gene expression data have been deposited in GEO: GSE76866.

SUPPLEMENTAL INFORMATION

Supplemental Information includes five figures and one table and can be found with this article online at <http://dx.doi.org/10.1016/j.immuni.2016.01.011>.

AUTHOR CONTRIBUTIONS

A.D.G. and M.C.N. conceived the study, interpreted the data, and wrote the manuscript. A.D.G. and Z.S. performed experiments. L.v.B. and A.G. performed antibody cloning and production. T.Y.O. performed bioinformatic analysis.

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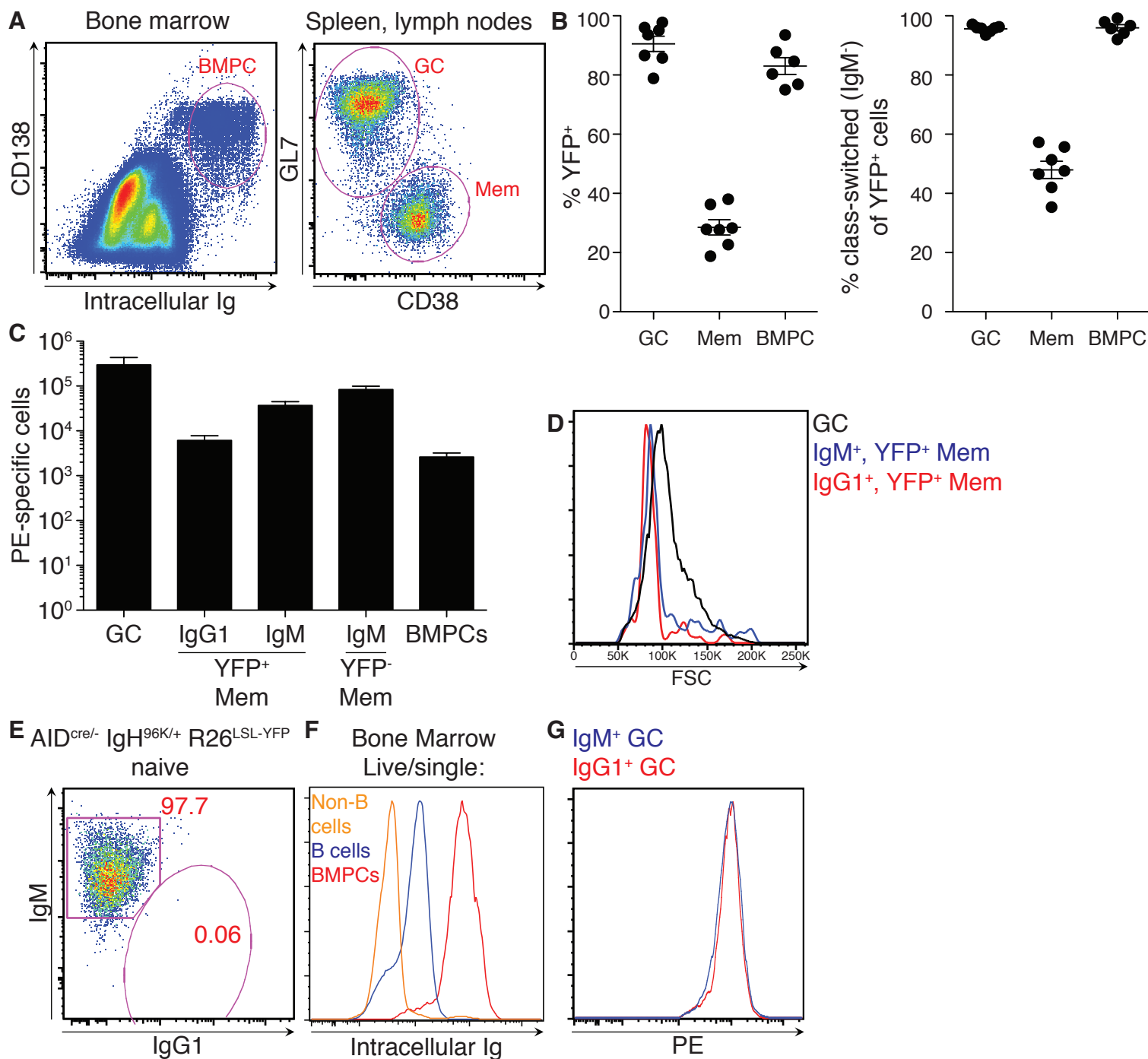
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Immunity, Volume 44

Supplemental Information

**Independent Roles of Switching
and Hypermutation in the Development
and Persistence of B Lymphocyte Memory**

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Figure S1**Figure S1. Characterization of antigen-specific B cells, related to Figure 2.**

(A) Live, single, and Dump⁻ cells were gated as shown (CD138⁺ intracellular Ig^{high}) to identify BMPCs after CD138-enrichment of bone marrow cells. GC and memory B cells were identified from spleen and lymph nodes based on CD38 and GL7 expression among antigen-specific B cells in *Aicda*^{cre/+} *R26*^{LSL-YFP} mice after immunization 32 days earlier with PE in CFA.

(B) Fraction of cells that are YFP⁺ (left panel) and class-switched (right panel) in indicated antigen-specific compartments of *Aicda*^{cre/+} *R26*^{LSL-YFP} mice, gated as shown in (A). Lines represent means.

(C) Average numbers of PE-specific cells enriched from spleen and lymph nodes or bone marrow of immunized *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP}.

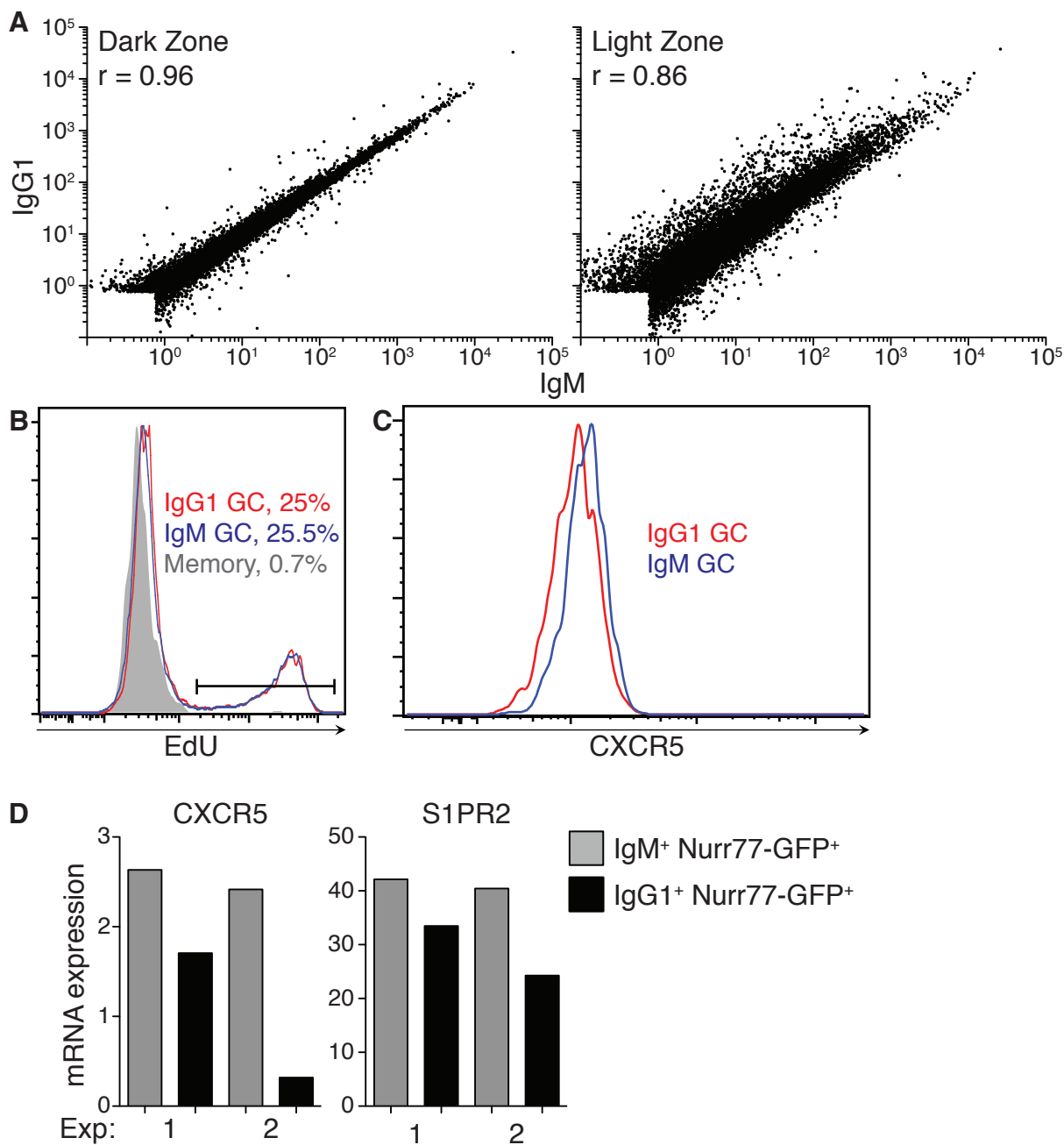
(D) Forward scatter plot of indicated PE-specific cell populations of immunized *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice.

(E) Isotype staining of PE-binding B cells in naive *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice.

(F) Intracellular Ig expression among indicated cell populations.

(G) PE-binding profiles of IgM⁺ and IgG1⁺ GC B cells in *Aicda*^{cre/-} *Igh*^{96K/+} mice immunized with PE in CFA 14 days earlier. Results are representative of one of 3 independent experiments.

Each dot represents one animal. Error bars = SEM.

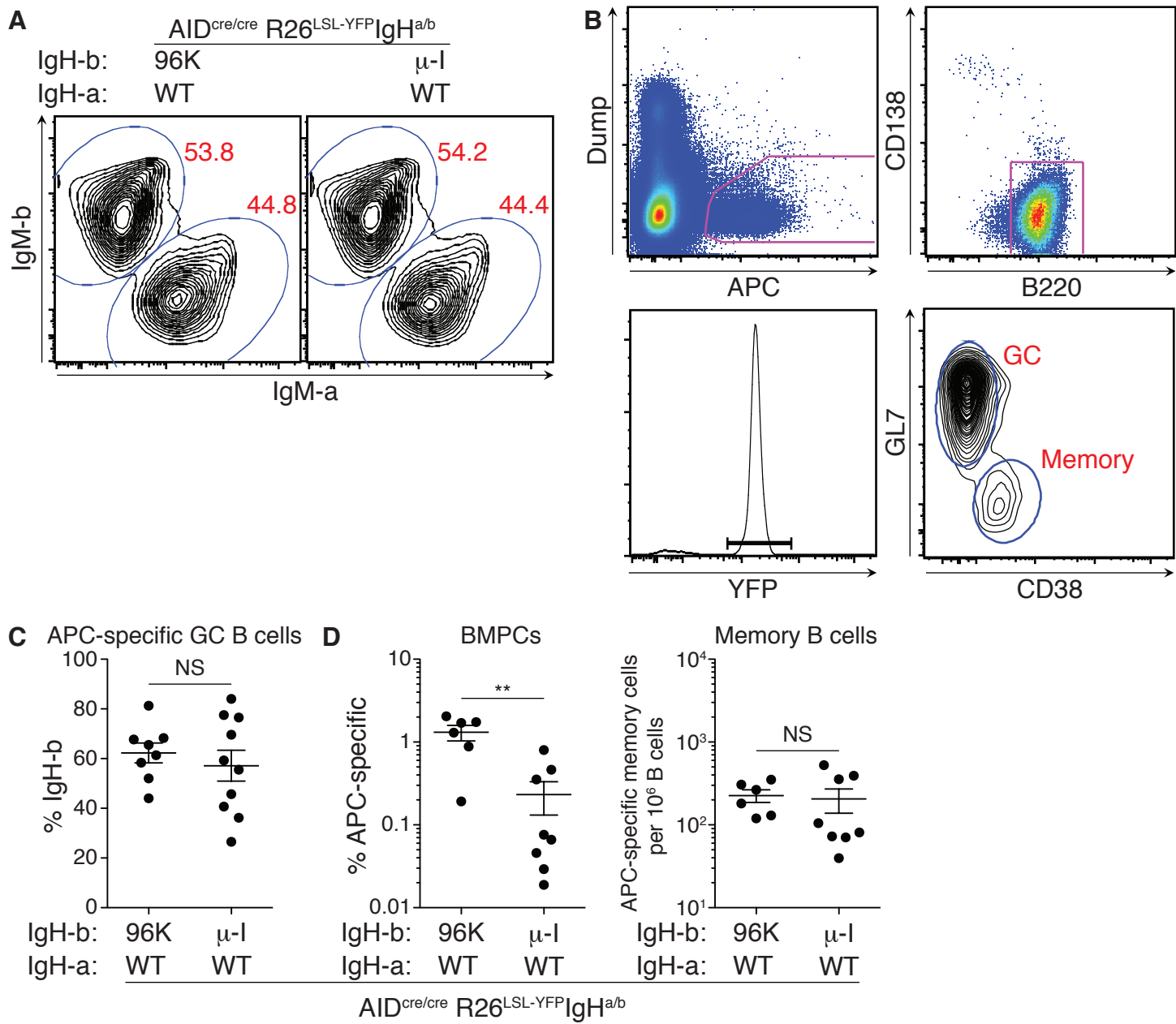
Figure S2**Figure S2. Characterization of IgG1⁺ and IgM⁺ GC cells, related to Figure 3.**

(A) RNA sequencing of IgG1⁺ and IgM⁺ light zone (CXCR4^{lo}CD86^{hi}) and dark zone (CXCR4^{hi}CD86^{lo}) cells generated by immunization of *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} with PE in CFA.

(B) EdU incorporation among PE-specific IgG1⁺ and IgM⁺ GC cells and memory B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice.

(C) CXCR5 expression among IgG1⁺ and IgM⁺ GC B cells in *Aicda*^{cre/-} *Igh*^{96K/+} *R26*^{LSL-YFP} mice immunized with PE in CFA.

(D) Quantitative PCR analysis of CXCR5 and S1PR2 transcripts relative to ACTB in IgG1⁺ and IgM⁺ Nurr77⁺ LZ cells of the *Aicda*^{cre/-} *Igh*^{96K/+} *Nurr77*-GFP⁺ genotype. Relative mRNA expression was multiplied by 10³ for clarity. All results represent two independent experiments.

Figure S3**Figure S3. Role of IgG1 in the GC in the absence of SHM, related to Figure 4.**

(A) IgM^a and IgM^b staining of naïve B cells (Dump⁺B220⁺CD38⁺GL7⁻YFP⁻) from mice of indicated genotypes.

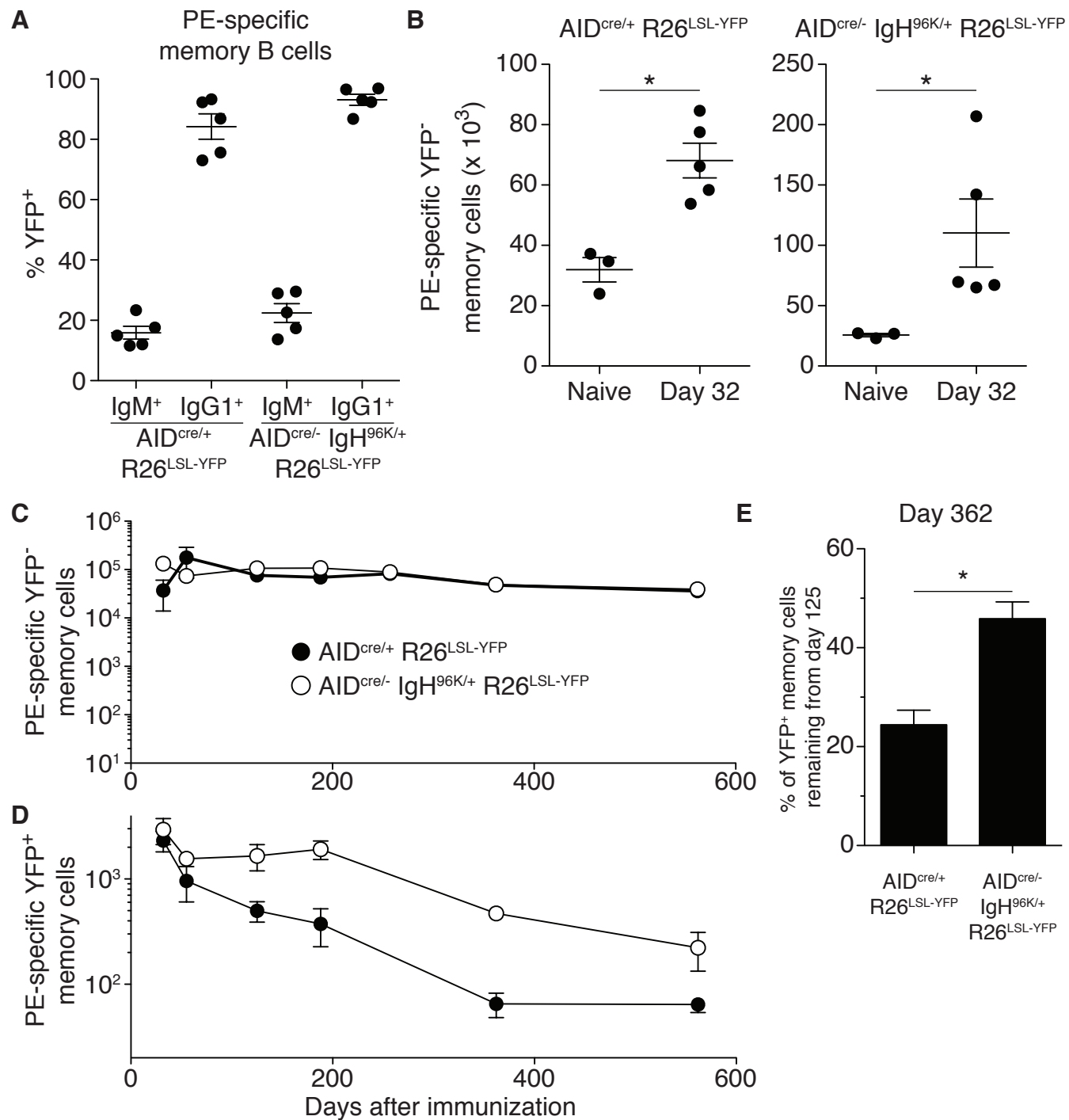
(B) Representative gating by flow cytometry of antigen-specific GC and memory B cells analyzed in Figures 4C-E.

(C) Fraction of antigen-specific YFP⁺ GC B cells expressing the IgH^b allotype in indicated mice 21 days after immunization with APC in CFA.

(D) Frequency of APC-specific BMPCs and number of APC-specific memory B cells per 10⁶ B cells generated in indicated mice 40 days after immunization with APC in CFA. Each dot represents one animal.

Lines represent means. Error bars = SEM. ** $p = 0.0047$.

Significance was assessed using two-tailed Mann-Whitney test. Data in (C) and (D) are combined from two independent experiments, as described in Figures 4A-E.

Figure S4**Figure S4. Characterization of memory cell populations and longevity, related to Figure 5.**

(A) Fraction of YFP⁺ cells among antigen-specific IgM⁺ and IgG1⁺ memory B cells of indicated mice 32 days after immunization.

(B) Number of YFP⁻ PE-specific memory B cells in indicated mice that were naïve or 32 days after immunization.

(C and D) Number of YFP⁻ (C) and YFP⁺ (D) PE-specific memory B cells in indicated mice over time after immunization with PE in CFA.

(E) Mean fraction of YFP⁺ PE-specific memory B cells from day 125 that remain at day 362 after immunization of indicated mice with PE in CFA. Fraction remaining was calculated as

(% YFP⁺ of PE-specific memory at day 362)/(% YFP⁺ of PE-specific memory at day 125) multiplied by 100.

* $p < 0.05$. Significance was assessed using a two-tailed Mann-Whitney test. Data are from two independent experiments. Each dot represents one animal. Error bars = SEM.

Figure S5

A

	VH	D	JH	CDR3	V _k	J _k	KLH	Insulin	ssDNA	dsDNA	LPS
EM10	IGHV1-81*01	IGHD1-1*01	IGHJ2*01	CARNGAIYYGSSFDYW	IGKV4-53*01	IGKJ1*01	-	-	-	-	-
EM13	IGHV1-81*01	IGHD1-1*02	IGHJ1*03	DGLYDRYFDVWG	IGKV5-43*01	IGKJ2*01	x	-	-	-	-
EM16	IGHV1-81*01	IGHD1-1*01	IGHJ4*01	WNYYGTLYPMDYWG	IGKV6-15*01	IGKJ2*01	x	x	x	x	-
EM19	IGHV1-81*01	IGHD2-3*01	IGHJ1*03	AYDGYPWYFDVWG	IGKV10-94*01	IGKJ1*01	-	-	-	-	-
EM26	IGHV1-62-2*01	IGHD2-4*01	IGHJ4*01	HEGHDYDGYAMDYWG	IGKV8-30*01	IGKJ1*01	-	-	-	-	-
EM28	IGHV1-81*01	IGHD1-1*01	IGHJ2*01	DNYYGSSGFDYWG	IGKV4-59*01	IGKJ1*01	-	-	-	-	-
EM3	IGHV1-78*01	IGHD2-13*01	IGHJ3*01	CARGRDGDAGNSRC	IGKV6-13*01	IGKJ5*01	x	x	x	x	x
EM30	IGHV1-81*01	IGHD2-5*01	IGHJ2*01	GSNYVGPFDYWG	IGKV4-53*01	IGKJ1*01	-	-	-	-	-
EM33	IGHV1-81*01	IGHD1-1*01	IGHJ2*01	SGYYGSSYFDYWG	IGKV4-53*01	IGKJ1*01	x	x	x	-	-
EM35	IGHV5-6*01	IGHD1-1*01	IGHJ2*01	RHYYGSDYFDYWG	IGKV4-53*01	IGKJ2*01	-	-	-	-	-
EM36	IGHV1-81*01	IGHD2-12*01	IGHJ2*01	TGSYYEYFDYWG	IGKV5-43*01	IGKJ1*01	x	x	x	x	x
EM39	IGHV1-81*01	IGHD1-1*01	IGHJ4*01	GDYYGSSSSGMDYWG	IGKV4-57*01	IGKJ4*01	-	-	-	-	-
EM40	IGHV5-6*02	IGHD1-1*01	IGHJ2*01	RHFYGSYFDYWG	IGKV4-53*01	IGKJ2*01	x	x	x	x	-
EM41	IGHV1-81*01	IGHD2-5*01	IGHJ2*01	TYYSNYDDYWG	IGKV4-57*01	IGKJ4*01	-	-	x	x	-
EM43	IGHV1-81*01	IGHD2-1*01	IGHJ3*01	VGYYGSAWCAHWG	IGKV10-94*01	IGKJ2*01	-	-	-	-	-
EM45	IGHV5-6*01	IGHD1-1*01	IGHJ2*01	RHYYGSDYFDYWG	IGKV4-53*01	IGKJ2*01	-	-	-	-	-
EM46	IGHV1-78*01	IGHD2-13*01	IGHJ2*01	QGRDYFDYWG	IGKV6-14*01	IGKJ5*01	-	-	-	-	-
EM49	IGHV1-76*01	IGHD3-2*02	IGHJ2*01	EQQLRPYYFDYWG	IGKV6-23*01	IGKJ4*01	-	-	-	-	-
EM5	IGHV1-81*01	IGHD2-5*01	IGHJ2*01	SNYGRFDCWG	IGKV14-100*01	IGKJ1*01	x	-	x	-	-
EM50	IGHV3-6*01	IGHD2-14*01	IGHJ1*03	RPDYYWYFDVWG	IGKV10-96*01	IGKJ2*01	-	-	-	-	-
EM51	IGHV1-81*01	IGHD3-2*02	IGHJ4*01	SSGYVGAMDYWG	IGKV4-53*01	IGKJ1*01	-	-	-	-	-
EM56	IGHV1-81*01	IGHD2-5*01	IGHJ2*01	GKDYYSKPFYWG	IGKV10-94*01	IGKJ2*01	x	x	x	x	-
EM57	IGHV1-72*01	IGHD2-10*01	IGHJ2*01	SLAIPYYFDYWG	IGKV6-13*01	IGKJ4*01	x	x	x	x	x
EM59	IGHV1-81*01	IGHD1-1*01	IGHJ4*01	GDYYGSSSSGMDYWG	IGKV4-57*01	IGKJ4*01	-	-	-	-	-
EM60	IGHV1-81*01	IGHD1-1*01	IGHJ1*03	SDYYGSSYWFYFDVWG	IGKV4-59*01	IGKJ5*01	-	-	-	-	-
EM61	IGHV1-81*01	IGHD1-1*01	IGHJ2*01	TNYDSSYPDYWG	IGKV10-94*01	IGKJ1*01	-	-	-	-	-
EM62	IGHV3-6*01	IGHD2-14*01	IGHJ1*03	RPDYYWYFDVWG	IGKV10-96*01	IGKJ2*01	-	-	-	-	-

B

	VH	D	JH	CDR3	V _k	J _k	KLH	Insulin	ssDNA	dsDNA	LPS
LM10	IGHV1-81*01	IGHD1-1*01	IGHJ4*01	SGYYGSSPMDYWG	IGKV4-53*01	IGKJ1*01	-	-	-	-	-
LM14-2	IGHV1-53*01	IGHD4-1*01	IGHJ2*01	SLENWATIYWGI	IGKV10-94*01	IGKJ5*01	-	-	-	-	-
LM15	IGHV1-85*01	IGHD1-1*01	IGHJ2*01	GLRYGFDYWG	IGKV10-94*01	IGKJ1*01	-	-	x	-	-
LM16	IGHV1-85*01	IGHD1-1*01	IGHJ2*01	GLRYGFDYWG	IGKV10-94*01	IGKJ1*01	-	-	-	-	-
LM17	IGHV1-81*01	IGHD4-1*01	IGHJ1*01	SGLGRGYFDVWG	IGKV12-44*01	IGKJ2*01	-	-	-	-	-
LM19	IGHV1-81*01	IGHD1-1*01	IGHJ4*01	SGHYGSSPLYAMDYWG	IGKV10-94*01	IGKJ2*01	-	-	-	-	-
LM2	IGHV1-9*01	IGHD1-1*01	IGHJ3*01	CARDYGKNYAWFPYWG	IGKV6-15*01	IGKJ2*01	-	-	-	-	-
LM21	IGHV1-53*01	IGHD2-1*01	IGHJ2*01	SLGISYGTIEDYWG	IGKV10-96*01	IGKJ1*01	-	-	-	-	-
LM22	IGHV1-55*01	IGHD2-5*01	IGHJ1*03	RRSSYSYNVNLWYFDVWG	IGKV1-117*01	IGKJ4*01	-	-	-	-	-
LM25	IGHV1-9*01	N/A	IGHJ4*01	HPYAMDYWG	IGKV10-94*01	IGKJ1*01	-	-	x	-	-
LM27	IGHV1-26*01	IGHD1-1*01	IGHJ4*01	RFLYGRVYAMDYWG	IGKV4-59*01	IGKJ2*01	-	-	-	-	-
LM3	IGHV1-81*01	IGHD1-1*01	IGHJ2*01	DYYADYFDYWG	IGKV9-120*01	IGKJ1*01	-	-	x	-	-
LM30	IGHV1-81*01	IGHD2-12*01	IGHJ2*01	NYYSDYEDYWG	IGKV9-120*01	IGKJ2*01	-	-	x	x	-
LM31	IGHV1-9*01	IGHD3-1*01	IGHJ4*01	KGYAMDYWG	IGKV10-94*01	IGKJ2*01	-	-	-	-	-
LM34	IGHV1-55*01	IGHD1-2*01	IGHJ4*01	SHYYDRRYKNNYAMDYWG	IGKV12-44*01	IGKJ2*01	x	-	-	-	-
LM37-2	IGHV1-78*01	IGHD2-3*01	IGHJ3*01	EEEYGYVRFAYWG	IGKV6-13*01	IGKJ1*01	-	-	-	-	-
LM38-2	IGHV1-55*01	IGHD1-1*01	IGHJ4*01	RFFFISTVVATGNYAMDSWG	IGKV1-117*01	IGKJ2*01	x	-	-	-	-
LM43-2	IGHV2-6*03	IGHD1-1*01	IGHJ4*01	HGGSRFDYAMDYWG	IGKV8-30*01	IGKJ5*01	-	-	-	-	-
LM52-2	IGHV1-59*01	IGHD5-7*01	IGHJ4*01	WRLDSAMDFWG	IGKV6-15*01	IGKJ4*01	-	-	-	-	-
LM55-2	IGHV1-81*01	IGHD2-3*01	IGHJ4*01	CARSGYGGMDYWG	IGKV4-59*01	IGKJ2*01	x	-	-	-	-
LM6	IGHV1-81*01	IGHD2-12*01	IGHJ2*01	SPPYNDYFDYWG	IGKV6-15*01	IGKJ2*01	-	-	x	-	-
LM60-2	IGHV1-55*01	IGHD1-1*01	IGHJ2*01	HYPGSGSSYFDYWG	IGKV1-117*01	IGKJ5*01	-	-	-	-	-
LM7	IGHV1-53*01	IGHD2-1*01	IGHJ2*01	SLGISYGTIEDYWG	IGKV10-96*01	IGKJ1*01	-	-	x	-	-
LM8	IGHV1-53*01	IGHD2-1*01	IGHJ2*01	SLGISYGTIEDYWG	IGKV10-96*01	IGKJ1*01	-	-	-	-	-
LM8-2	IGHV1-53*01	IGHD4-1*01	IGHJ2*01	SLENWATIYWG	IGKV10-94*01	IGKJ5*01	-	-	-	-	-

Figure S5. Early and late memory antibodies, related to Figure 6. Repertoire and reactivity of early (EM) and late (LM) memory antibodies.

Table S1. Gene expression among Nurr77-GFP⁺ GC B cells, related to Figure 3.

List of genes differentially expressed by a fold-change of at least 0.6 (\log_2) among Nurr77-GFP⁺ IgM⁺ and IgG1⁺ LZ cells.